



TITLE OF THE INVENTION

DECREASED FAT ABSORPTION WITH AN ANTI-LIPASE ANTIBODY

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BACKGROUND OF THE INVENTION

Field of the invention

A food additive that decrease fat absorption in mammals

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Discussion of the background

Our sedentary life including the decreased physical activity and increased food intake have made us prone to be overweight. The above has brought as consequence that almost 40-50 % of the USA population is 20% above their desirable weight. The advance in the science of food and nutrition not only has made us wiser about the functions of all nutrients but also by applying that knowledge we have concentrated food in smaller portions by which the satisfaction of filling is decreased. Even if the amount of food intake remains the same, we will have an excess caloric intake due to the high energy concentration of such type of food (Bell, et al 1997). Currently, the weight loss related market is full of diet

pills that reduce appetite by suppressing brain hormones, drugs that suppress the absorption of nutrients, pills that supposedly have ergogenics effect, pills that increase food passage rate and other fad diets. Mostly all of these drugs
5 have secondary effects like depression, anxiety, addiction and others.

A new approach for the reduction of calories in food is by the use of fat substitutes (Gershoff, et al 1995). Each gram of fat provides 9 calories as compared to 4 calories per
10 gram of carbohydrate and protein. Fat substitutes mainly those made of long carbohydrate chains are use for the elaboration of prepared food with the purpose of maintaining fat properties in the prepared food but decreasing calories. A new fat substitute, Olestra, which is made of long chain
15 fatty acids that are too big for digestive enzymes (lipase) to breakdown, therefore that type of fat is not absorbed. It has been observed that the consumption of Olestra has resulted in decreased absorption of fat soluble and the presence of fat in the feces. A long term study (12 weeks)
20 where 1/3 of the dietary fat was replaced with olestra, female subjects lost weight and did not compensate for the reduced calories and fat intake (Roy, et al, 1997).

In the animal industry, researchers have been working on the reduction of fat accumulation in animals since this characteristic first, has a negative effect on profits and second, consumers want less visible fat in order to decrease the health risk.

Fat accumulation in animals has been reduced by passively administered antibodies against adipocyte plasma membrane in rats, pigs, rabbits and lambs. Immunity against growth hormone has also decreased abdominal fat in chickens (Brodie and Hu, 1996; Moloney, 1995; Flint, 1992).

Lipase, an enzyme produced by the pancreas, hydrolyzes triacylglycerides into free fatty acids and glycerol. This is a crucial step in breaking down ingested fat in the gastrointestinal tract. Lipase is more active in the duodenum (small intestine) where broken down fat with the aid of bile salts form micelles and then are absorbed by the intestinal mucosa.

Therefore, by inhibiting lipase the ingested fat will not be absorbed and the energy supplied by fat and the fat itself will be excreted.

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DESCRIPTION OF THE PREFERRED EMBODIMENTS

20 Previous research on the effectiveness of chicken
antibodies has been reported; i.e. the prevention of
bacterial infection in swine, calf and dairy cows (Yokoyama
et al, 1993; Erhard et al 1993; Coleman, 1995). These

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researches have also demonstrated the presence of intact avian antibodies in the gastro-intestinal tract of the animals.

Although chickens antibodies are known to protect
5 against bacterial infections, no antibody has been reported to decreased fat absorption.

It will be apparent for those skilled in the art that the aforementioned objects and other advantages may be further achieved by the practice of the present invention.

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Example 1

This example illustrate the preparation of the specific antibody against lipase. 17-week old hens were injected with 2.5 mg of lipase (Sigma Chemical Co.). The inoculum was
15 prepared by dissolving the enzyme in 0.2 ml phosphate buffered saline (PBS, pH 7.3) and 0.2 ml complete Freund's adjuvant. The antigen preparation was injected into two sites 0.2 ml in each (right and left) pectoralis muscle. A total of 0.4 ml of antigen preparation per hen was administered. A
20 second injection was administered 5-6 weeks following the initial injection (at about 50% egg hen production). In the second antigen preparation, incomplete Freund's adjuvant was used instead of complete Freund's adjuvant. Hens were re-

injected with the antigen preparation every two months or when the antibody titer was determined to be low. Antibody titer was determined by ELISA. Hens had free access to feed and water and they were maintained in an isolated room in order to minimize outside contamination.

Example 2

Antibody was purified as follows: One volume of egg yolk of example 1 was mixed with 9 volumes of distilled water and left to sit overnight at 4 °C. Then the aqueous portion was centrifuged at 4000 rpm for 10 minutes and filtered through a cheesecloth in order to remove any excess fat. The aqueous portion contains all the protein present in the egg yolk which includes all the antibodies (IgY). The liquid was frozen and then was freeze dried. The antibody activity was determined by ELISA.

Example 3

Antibody against lipase was determined as follows:

1.- ELISA plates were coated with 100 ul lipase preparation (50 ug/ml) in carbonate buffer. The plates were incubated at 4 °C overnight prior to blocking with 1.5% bovine serum albumin for 4 hours at room temperature.

2.- 100 ul of a 0.5 mg protein/ml antibody extract was added to each well and the plates incubated at room temperature for 1 hour.

3.- Plates were washed with PBS-tween solution. 100 ul of rabbit anti-chicken IgG conjugated to horseradish peroxidase was added to each well. The plates were incubated at room temperature for 1 hour.

4.- Plates were washed with PBS-tween and 100 ul of TMB substrate was added to each well and incubated for 15 minutes.

5.- The reaction was stopped with 100 ul of 2 M sulfuric acid.

6.- Plates were read at 455 nm in an ELISA plate reader.

7.- Titer was determine as the inverse of the dilution at which O.D. of the immunized egg was similar to the un-immunized control (O.D. < 0.100).

Example 4

This study illustrates the in vitro inhibition of lipase by the chicken anti-lipase antibody. The effectiveness of the antibody was verified by using a test specific for the determination of lipase in serum (Sigma Chemical Co). We modified this test by adding a known amount of enzyme

(lipase) and antibody to a certain volume of phosphate buffered saline. The resulting activity was expressed as Sigma-Tietz units/ml, which is equal to the ml of 0.05 N NaOH required to neutralize the fatty acid formed in the reaction.

5 In a preliminary study we found the following:

| Lipase (mg) | anti-lipase (protein extract) (mg) | Lipase Activity (U) | % decreased activity |
|----------------|--|---------------------------|----------------------------|
| 2.0 | 0 | 17.3 | |
| 2.0 | 9.0 | 18.8 | 0 |
| 1.0 | 0 | 14.0 | |
| 1.0 | 9.0 | 12.5 | 11 |
| 0.5 | 0 | 10.4 | |
| 0.5 | 9.0 | 9.8 | 6 |
| 0.25 | 0 | 8.1 | |
| 0.25 | 9.0 | 6.7 | 17 |

In a second test; higher amount of antibody extract was used. The results are as follows:

| Lipase (mg) | Anti-lipase Protein Extract (mg) | Lipase Activity (U) | % decreased activity |
|----------------|---|---------------------------|-------------------------|
| 2.0 | 0 | 18.7 | |
| 2.0 | 37 | 14.0 | 25 |
| 1.0 | 0 | 13.5 | |
| 1.0 | 37 | 6.9 | 49 |

Example 5

This study illustrates the effect of anti-lipase antibody in mice. ~~Two groups of 5 2-month old mice (25-34 gr each)~~ were given 5 mg of antibody (protein extract) per ml of water. The antibody was mixed with water on a daily basis. Mice were fed the same amount of feed in both groups (approx. ~~5 gr/mice/day~~). The length of the experiment was 58 days. The results are as follows

| | total* initial body weight (gr) | total final body weight (gr) | difference in body weight (gr) | total feed intake (gr) | gr of feed needed to gain 1 gr of body weight |
|-------------|---|--|---|---------------------------------|--|
| control | 157 | 199 | 42 | 1039 | 24.74 |
| anti-lipase | 156 | 187 | 31 | 1039 | 33.52 |

* Sum of 5 mice/trt

Example 6

This study illustrate the effect of anti-lipase when fed at lower levels than in example 5. Two groups of 5 5-month old mice (32-35 gr each) were given 1 mg of antibody (protein extract) per ml of water for the first 7 days then it was increased to 2 mg/ml of water. Mice in both groups were fed the same amount of feed for 35 days. The results were as follows.

| | Total initial body weight (gr) | Total final body weight (gr) | difference in body weight | total feed intake (gr) | gr of feed needed to gain 1 gr of body weight |
|-------------|--|--|---------------------------------|---------------------------------|--|
| control | 182 | 204 | 22 | 787.4 | 35.79 |
| anti-lipase | 181 | 185 | 4 | 788.5 | 197.13 |

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Example 7

This study demonstrate the encapsulation of anti-lipase antibodies by liposomes. This liposome preparation was based on the procedure by Shimizu et al (1993). Final liposome suspension was frozen and later freeze dried. A known amount

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of freeze dried liposome was mixed with rat diet, and fed daily for the length of the study.

Example 8

5 This study illustrate the effect of anti-lipase antibodies in rats. Twelve retired breeder Sprague Dawley rats (Harlan, Wisconsin) were individually caged and supplied with free access to water. They were fed a rabbit chow which was supplemented with corn oil in order increase fat content
10 to 30%. Feed intake was monitored for 1 week in order to determine the amount of feed needed to maintain their initial body weight. Rats were divided in two groups one fed the high fat diet and the other group was fed the same diet with freeze dried liposome containing anti-lipase antibody
15 extract. The treated diet contained 750 mg antibody/kg of diet. The results after 1 week of treatment are as follows:

| | Initial body weight (gr) * | One week feed intake (gr) | Final body weight (gr) | grams of feed needed to gain 1 gr of body weight |
|----------|----------------------------------|---------------------------------|------------------------------|--|
| control | 316 | 132.3 | 327 | 12.0 |
| antibody | 319 | 129.4 | 326 | 18.5 |

* average of 6 rats.

Example 9

Since it was observed that rats gained weight in example 8, the same rats were used but this time feed was restricted.

5 The results are as follows:

| | initial body weight (gr) | Final body weight (gr) | difference in body weight | feed intake (gr) |
|----------|--------------------------------|------------------------------|---------------------------------|------------------------|
| control | 327 | 319 | -8 | 102 |
| antibody | 326 | 317 | -9 | 101 |

Example 10

10 This study demonstrate the effect of the anti-lipase when fed to rats at maintenance feed intake. The results are as follows:

| | initial body weight (gr) | One week body weight (gr) | difference in body weight | One week feed intake (gr) | gr of feed needed to gain 1 gr of body weight |
|----------|-----------------------------------|---------------------------------------|---------------------------------|---------------------------------------|---|
| control | 325 | 332 | 7 | 112 | 16 |
| antibody | 324 | 325 | 1 | 114 | 114 |

It will be apparent to those skilled in the art that a number of modifications and variations may be made without departing from the scope of the present invention as set forth in the appended claims.

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